



The Next Generation in Carbon Sequestration Solutions

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Relating to Section 5 on Offsets:
National Emissions Trading Taskforce Discussion Paper

An accurate and quantifiable method of terrestrial carbon sequestration for Australian Farmers, Landholders and other Land-Managers

Introduction

Australia has 20 million hectares under cultivation of grass crops annually, salinity affects a further 2.5 million hectares and grazing land exceeds both of these areas combined. Phytolith (or Plantstone) carbon sequestration is relevant to a wide range of plant types including those grown for agricultural crop production, grazing and the rehabilitation of degraded landscapes. The following method of offsets through Phytolith organic carbon (**PhytOC**) sequestration allows farmers **for the first time** to be included and **rewarded** for their efforts in **carbon abatement**. Significantly this may be achieved **accurately** without costly or approximate calculation processes required for monitoring soil carbon stocks themselves. Nevertheless, the contribution of soil carbon stocks does need to be recognised and introduced into any offset scheme perhaps using no-till calculations excepted under some carbon abatement schemes elsewhere. The **PhytOC** process involves a land-use change or, the intentional selection and replacement of one species or variety of a crop/plant to another for which the additionally of carbon sequestered can be scientifically verified and measured with a high degree of accuracy. Baseline data is accurately measured and easily monitored using the method proposed below. The concept is as follows:

The baseline measurement is taken for the existing plant type or the variety you would normally sow e.g. a wheat variety with e.g. a **Low** PhytOC sequestration capacity.

The additionality is achieved by selecting and then planting/sowing a plant type or variety e.g. a wheat variety with **High** PhytOC sequestration capacity.

The Verification is made by (1) using archived baseline data on the PhytOC storage capacity of specific varieties that we are currently making available or, (2) by sending samples of both existing and new plants/varieties to be tested at a suitably accredited laboratory at minimal cost to the producer/landholder using published methods (c.f. Parr and Sullivan 2005) and a standard carbon analyzer, where:

$$H - L = A$$

High % of PhytOC capacity (H), minus the **Low** % PhytOC capacity (L), equals the **Additional** carbon sequestered (A)

This method of PhytOC sequestration is compatible with existing guiding principles of good practice in a number of sections of the IPCC Guidelines c.f. section 3.1.2.3 “Other Managed Lands” and as footnoted under these guidelines “*National circumstances may necessitate slight **modifications to the pool definitions** used here. Where modified definitions are used, it is good practice to report upon them clearly, to ensure that modified definitions are used consistently over time, and to demonstrate that pools are **neither omitted** nor double counted*”.

Thus it can be interpreted from this statement that while it is very important that we stick to the intended principles of Kyoto and good practice guidelines under IPCC, the definitions themselves should be viewed as a guide rather than an absolute in order that important carbon pools not be excluded.

Keeping this in mind, in Chapter 5 of the National Emissions Trading Taskforce Discussion Paper dealing with offsets the main requirements set out are:

Consistency with international frameworks

NETS p66. It is proposed that approaches to facilitate the development of offset projects and the creation of offset credits under the NETS be as consistent as possible with those emerging methodologies, rules and frameworks being developed for the JI mechanism under the Kyoto Protocol.

As pointed out above the method of PhytOC sequestration is consistent with the principals of the Kyoto Protocol and compatible with existing guiding principles of good practice in a number of sections of the IPCC Guidelines.

Additionality

NETS p66-67. The key aim of an offsets regime is to provide an incentive for abatement that would not otherwise have occurred... This means that it is possible to identify what would be the most likely activity under the 'business as usual' scenario in a variety of sectors.

The **PhytOC** sequestration process requires a definite change from a business as usual scenario by the necessity to make a conscious decision to shift from the practice of growing a **Low** % PhytOC capacity variety or plant type (and in the case of e.g. degraded saline soils a **no** % PhytOC capacity) to intentionally introducing a plant type or variety with a **High** % of PhytOC capacity.

Baseline setting and monitoring

Along with published data an independent report outlining in more detail than can be provided here, on how the baseline setting and monitoring could be implemented for PhytOC was submitted to the NSW Greenhouse Abatement Scheme Administrator and IPART by the Australian Forest Corporation an accredited NSW Greenhouse Abatement Certificate Provider. However, briefly as outlined above the baseline measurement is easily taken for the existing plant type or the variety you would normally sow. This information might be retrieved from archived information on the PhytOC capacity or by submitting the sample to a suitably qualified laboratory and assessed using published internationally excepted standard methods of carbon (PhytOC) quantification (c.f. Parr and Sullivan 2005).

Field trials

The annually reported yields in the fodder, grain and other varieties that we have found to date to contain high PhytOC have been equal to or up with the best yeilders for the season particularly for sugarcane and sorghums. Thus there is no need to experience a loss in yield or mass for the land manager simply a decision to improve carbon sequestration rates by selecting a high **PhytOC** capacity variety.

For example in sugarcane field trials in northeastern NSW the 'additional' soil carbon sequestration rate for the high PhytOC yielding sugar cane variety over the low PhytOC yielding sugar cane varieties was the equivalent of ~ 0.4 tonne eCO_2 $ha^{-1}yr^{-1}$. In field trials on sorghum at Tamworth NSW we had seven varieties growing in the same soil and, of those varieties two were found to have exceptionally high **PhytOC** levels. The best of these had a **PhytOC** capacity ten times that of the average variety grown and was reported in one season to have the highest harvest yields at two sites in Queensland.

Note: The PhytOC sequestering method may be used in conjunction with other proposed procedures for carbon abatement such as increased ground cover and no-till farming. For agricultural crops the PhytOC component that is measured is in the post-harvest stubble thus there is no loss in grain production etc.

A brief explanation of the science behind the PhytOC carbon sequestration concept is as follows:

(a) All grasses contain silica Phytoliths (or Plantstones) in their outer (epidermal) cells, this is a natural plant trait (Bozarth, 1992; Brown, 1984a; Brown, 1984b; Clifford and Watson, 1977; Fredlund and Tieszen, 1997a; Krishnan et al., 2000; Krull et al., 2003; Lentfer et al., 1997; Mulholland and Rapp, 1992b; Ollendorf et al., 1988; Parr, 2004; Parr and Sullivan, 2005b; Piperno and Pearsall, 1998; Thorn, 2004; Twiss, 1992; Twiss et al., 1969).

(b) The silica entombs cellular material comprised of carbon that has been identified using a number of methods (Allmaras and Albrecht, 2006; Boyd and Lentfer, 1995; Fredlund and Tieszen, 1997b; Golchin et al., 1994; Houyuan. et al., 2000; Kelly et al., 1991; Krull et al., 2003; Parr and Sullivan, 2005b; Piperno, 1998; Wilding, 1967; Wilding et al., 1967).

(c) When a plant dies and decays or, it is harvested and the stubble is left in the paddock, the silica Phytoliths are incorporated into the soil matrix c.f. (Allmaras and Albrecht, 2006; BOWMAN et al., 2004; Boyd and Lentfer, 1995; Fredlund and Tieszen, 1997b; Golchin et al., 1994; Houyuan. et al., 2000; Kelly et al., 1991; Krull et al., 2003; Parr and Sullivan, 2005b; Piperno, 1998; Wilding, 1967; Wilding et al., 1967).

(c) Silica Phytoliths are durable and can take thousands of years to brake down in soils (Albert et al., 2000; Allmaras and Albrecht, 2006; Bowdery, 1989; Bowdery, 1996; Boyd, 1999; Boyd et al., 2005; Boyd et al., 1998b; Kealhofer and Piperno, 1994; Parr, 2003; Parr and Carter, 2003; Parr et al., 2001; Pearsall, 1989; Pearsall, 1999; Pearsall and Trimble, 1984; Piperno, 1994; Piperno, 1985; Piperno, 1998; Piperno et al., 2000).

(d) This carbon entombed in silica Phytolith cells is, as a result, sequestered for the long-term demonstrated by radiocarbon dating the Phytoliths themselves (c.f. 8,000-12,000 years (Mulholland and Prior, 1993; Parr and Sullivan, 2005a; Parr and Sullivan, 2005b; Sullivan and Parr, 2005; Wilding, 1967; Wilding et al., 1967)

(e) The amount of carbon locked up by each species and/or variety of a crop or grass, varies significantly. Thus, if you choose a higher yielding PhytOC variety over a lower yielding variety you presumably fulfill the additionally clause requirement by the IPCC (Kyoto) guidelines for carbon abatement (Parr, 2003; Parr and Sullivan, 2005a; Parr and Sullivan, 2005b; Sullivan and Parr, 2005).

(f) The beauty of this method is that you only need to test the plant varieties for the baseline PhytOC content rather than the soils themselves, which is cheap, easy, quick and very accurate.

(g) Unlike other carbon fractions currently approved and measured PhytOC carbon is resistant to decomposition, fire and oxidation under most natural conditions.

References Cited:

Albert, R.M., Weiner, S., Bar-Yosef, O. and Meignen, L., 2000. Phytoliths in the middle palaeolithic deposits of Kebara Cave, Mt Carmel, Israel: study of the plant materials used for fuel and other purposes. Journal of Archaeological Science, 27: 931 - 947.

Allmaras, R. and Albrecht, S., 2006. Occluded C in Phytoliths: a Potential Mechanism for Carbon Sequestration in a Pacific Northwest Mollisol., 18th World Congress of Soil Science, Philadelphia.

Bowdery, D., 1989. Phytolith analysis: introduction and applications., In: W. In. Beck, Clarke, A. & Head, L. (Editor), *Plants in Australian Archaeology*, Archaeology and Material Culture Studies in Anthropology. Watson Ferguson & Company., Brisbane., pp. Tempus Vol. (1):161 - 86.

Bowdery, D.E., 1996. Phytolith analysis applied to Archaeological Sites in the Australian arid zone. Unpublished PhD Thesis, Unpublished PhD Thesis, Australian National University, Canberra.

Bowman, D.M.J.S., Cook, G.D. and Zoppi, U., 2004. Holocene boundary dynamics of a northern Australian monsoon rainforest patch inferred from isotopic analysis of carbon., *Austral Ecol*, 29(6): 605-612.

Boyd, W.E., 1999. The geoarchaeology of Numondo, W.N.B. (P.N.G.): Late Pleistocene and Holocene landscape evolution.

Boyd, W.E. and Lentfer, C.J., 1995. Dating Fossil Phytoliths. *AINSE Activities*, 10: 7.

- Boyd, W.E., Lentfer, C.J. and Parr, J.F., 2005. Interactions between human activity, volcanic eruptions and vegetation during the Holocene at Garua and Numundo, West New Britain, PNG. *Quaternary Research*, 64: 384-398.
- Boyd, W.E., Lentfer, C.J. and Torrence, R., 1998b. Phytolith analysis for a wet tropics environment: methodological issues and implications for the archaeology of Garua Island, West New Britain, Papua New Guinea. *Palynology*, 22: 213 - 228.
- Bozarth, S.R., 1992. Classification of opal phytoliths formed in selected dicotyledons native to the Great Plains. In: J. George Rapp and S.C. Mulholland (Editors), *Phytolith Systematics Emerging Issues*. Society for Archaeological Sciences, New York.
- Brown, D.A., 1984a. Prospects and limits of a phytolith key for grasses in the central United States. *Journal of Archaeological Science*, 11: 345 - 68.
- Brown, D.A., 1984b. Prospects and limits of phytolith key for grasses in the Central United States. *Journal of Archaeological Science*, 11: 345-68.
- Clifford, H.T. and Watson, L., 1977. *Identifying Grasses: Data, Methods and Illustrations*. University of Queensland Press, Brisbane.
- Fredlund, G.G. and Tieszen, L.L., 1997a. Calibrating grass phytolith assemblages in climatic terms: Application to late Pleistocene assemblages from Kansas and Nebraska. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 136: 199-211.
- Fredlund, G.G. and Tieszen, L.L., 1997b. Phytolith and Carbon Isotope Evidence for Late Quaternary Vegetation and Climate Change in the Southern Black Hills, South Dakota. *Quaternary Research*, 47: 206-217.
- Golchin, A., Oades, J.M., Skjemstad, J.O. and Clarke, P., 1994. Study of Free and Occluded Particulate Organic Matter in Soils by Solid-state ¹³C CP/MAS NMR Spectroscopy and Scanning Electron Microscopy. *Australian Journal of Soil Science*, 32: 285-309.
- Houyuan., L., Yongji., W., Guo'an., W., Hui., Y. and Zhen., L., 2000. Analysis of carbon isotope in phytoliths from C3 and C4 plants and modern soils. *Chinese Science Bulletin*, 45(19): 1804-1807.
- Kealhofer, L. and Piperno, R.P., 1994. Early agriculture in southeast Asia: phytolith evidence from the Bang Pakong Valley, Thailand. *Antiquity*, 68: 564-572.
- Kelly, E.F., Amundson, R.G., Marino, B.D. and Deniro, M.J., 1991. Stable isotope ratios of carbon in phytoliths as a quantitative method of monitoring vegetation and climate change. *Quaternary Research*, 35: 222 - 233.
- Krishnan, S., Samson, N.P., Ravichandran, P., Narasimhan, D. and Dayanandan, P., 2000. Phytoliths of Indian grasses and their potential use in identification. *Botanical Journal of the Linnean Society*, 132: 241 - 252.
- Krull, E.S. et al., 2003. ¹³C-depleted charcoal from C3 and C4 grasses and the role of occluded carbon in phytoliths. *Organic Geochemistry*, 34: 1337-1352.
- Lentfer, C.J., Boyd, W.E. and Gojak, D., 1997. Hope Farm Windmill: phytolith analysis of cereals in early colonial Australia. *Journal of Archaeological Science*, 24: 841 - 856.
- Mulholland, S.C. and Prior, C.A., 1993. AMS radiocarbon dating of phytoliths. In: D.M. Pearsall and D.R. Piperno (Editors), *MASCA Research Papers in Science and Archaeology*. University of Pennsylvania, Philadelphia, pp. 21 - 23.
- Mulholland, S.C. and Rapp, G.J., 1992b. A Morphological Classification of Grass Silica Bodies. In: G.R.J.a.S.C. Mulholland (Editor), *Phytolith Systematics: Emerging Issues*. Plenum Press, New York, pp. 65-89.
- Ollendorf, A.L., Mulholland, S.C. and Rapp, J.G., 1988. Phytolith analysis as a means of plant identification: *Arundo donax* and *Phragmites communis* *Annals of Botany*, 61: 209 - 214.
- Parr, J.F., 2003. A study of Palaeo-Landscapes in the Numundo region of West New Britain, Papua New Guinea, as indicated by Fossil Phytolith Analysis. Unpublished PhD. Thesis, Unpublished PhD. Thesis, Southern Cross University.
- Parr, J.F., 2004. Morphometric and visual fossil phytolith identification using a regionally specific digital database. *Phytolitharian*, 16(2): 2-10.
- Parr, J.F. and Carter, M., 2003. Phytolith and starch analysis of sediment samples from two archaeological sites on Dauar Island, Torres Strait. *Vegetation History and Archaeobotany*, 12(2): 131-141.
- Parr, J.F., Lentfer, C.J. and Boyd, W.E., 2001. Spatial analysis of phytolith assemblages at an archaeological site in West New Britain, Papua New Guinea. In: G.R. Clark, A.J. Anderson and T. Vunidilo. (Editors), 'The

Archaeology of Lapita Dispersal in Oceania'. Pandanus Press,, Australian National University, Terra Australis, pp. (17) pp. 125-134.

- Parr, J.F. and Sullivan, L.A., 2005a. Carbon Sequestration in Plantstones. In: C. Jones (Editor), Managing the Carbon Cycle: Forum, Armidale, pp. 23-28.
- Parr, J.F. and Sullivan, L.A., 2005b. Soil carbon sequestration in Phytoliths. *Soil Biology and Biochemistry*, 37(1): 117-124.
- Pearsall, D.M., 1989. *Paleoethnobotany: A Handbook of Procedures*., Academic Press, Inc., London.
- Pearsall, D.M., 1999. Agricultural evolution and the emergence of formative societies in Ecuador. In: M. Blake (Editor), *Pacific Latin America in Prehistory: The Evolution of Archaic and Formative Cultures*. Washington State University Press, Washington, pp. 161 - 170.
- Pearsall, D.M. and Trimble, M.K., 1984. Identifying past agricultural activity through soil phytolith analysis: a case study from the Hawaiian Islands. *Journal of Archaeological Science*, 11: 119 - 133.
- Piperno, D., 1994. Phytolith and charcoal evidence for prehistoric slash-and-burn agriculture in the Darien rainforest of Panama. *The Holocene*, 4(3): 321-325.
- Piperno, D.R., 1985. Phytolith analysis and tropical paleo-ecology: production and taxonomic significance of siliceous forms in new world plant domesticates and wild species. *Review of Palaeobotany and Palynology*, 45: 185 - 228.
- Piperno, D.R., 1998. Paleoethnobotany in the Neotropics from microfossils: new insights into ancient plant use and agricultural origins in the tropical forest. *Journal of World Prehistory*, 12(4): 393 - 449.
- Piperno, D.R., Andres, T.C. and Stothert, K.E., 2000. Phytoliths in *Cucurbita* and other Neotropical Cucurbitaceae and their occurrence in early archaeological sites from the lowland American tropics. *Journal of Archaeological Science*, 27: 193 - 208.
- Piperno, D.R. and Pearsall, D.M., 1998. The silica bodies of tropical American grasses: morphology, taxonomy, and implications for grass systematics and fossil phytolith identification. *Smithsonian Contributions to Botany*, 85: 40.
- Sullivan, L.A. and Parr, J.F., 2005. The potential of soil to Securely Sequester carbon: expanding the horizon. In: C. Jones (Editor), *Managing the Carbon Cycle: Forum*, Armidale, pp. 17-22.
- Thorn, V.C., 2004. Phytolith evidence for C4-dominated grassland since the Holocene at Long Pocket, northeast Queensland, Australia. *Quaternary Research*, 61: 168-180.
- Twiss, P.C., 1992. Predicted World Distribution of C3 and C4 Grass Phytoliths., In: G.R.J.a.S.C. Mulholland (Editor), *Phytolith Systematics: Emerging Issues*. Plenum Press, New York, pp. 113-128.
- Twiss, P.C., Suess, E. and Smith, R.M., 1969. Morphological classification of grass phytoliths. *Soil Science Society of America Proceedings*, 33: 109 - 115.
- Wilding, L.P., 1967. Radiocarbon dating of biogenetic opal. *Science*, 156: 66 - 67.
- Wilding, L.P., Brown, R.E. and Holowaychuk, N., 1967. Accessibility and properties of occluded carbon in biogenetic opal. *Soil Science*, 103(1): 56 - 61.
- Wilding, L.P. and Drees, L.R., 1974. Contributions of forest opal and associated crystalline phases to fine silt and clay fractions of soils. *Clay and Clay Minerals*, 22: 295-306.